

USING ASPARTYL NITROBENZOTHAZINE TO IMPROVE GLYCOLIGATION STRATEGY

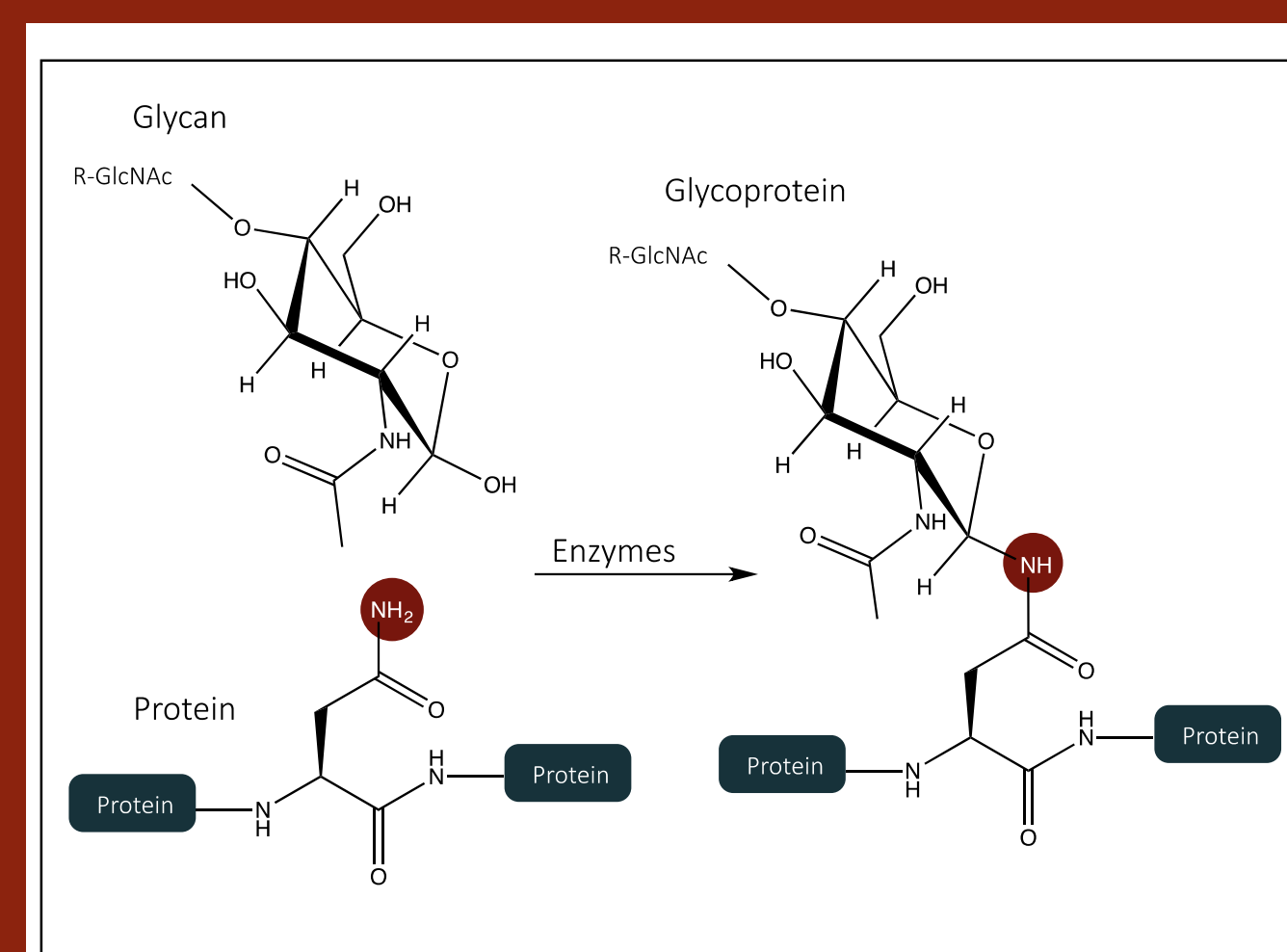
¹Tyler Siegford, ²Daniel Collins, ²Philip Garner, and ³Darren Thompson (mentor)

UNIVERSITY OF IDAHO¹, WASHINGTON STATE UNIVERSITY², UNIVERSITY OF IDAHO COEUR D'ALENE – PEPTIDAHO RESEARCH CONSORTIUM³

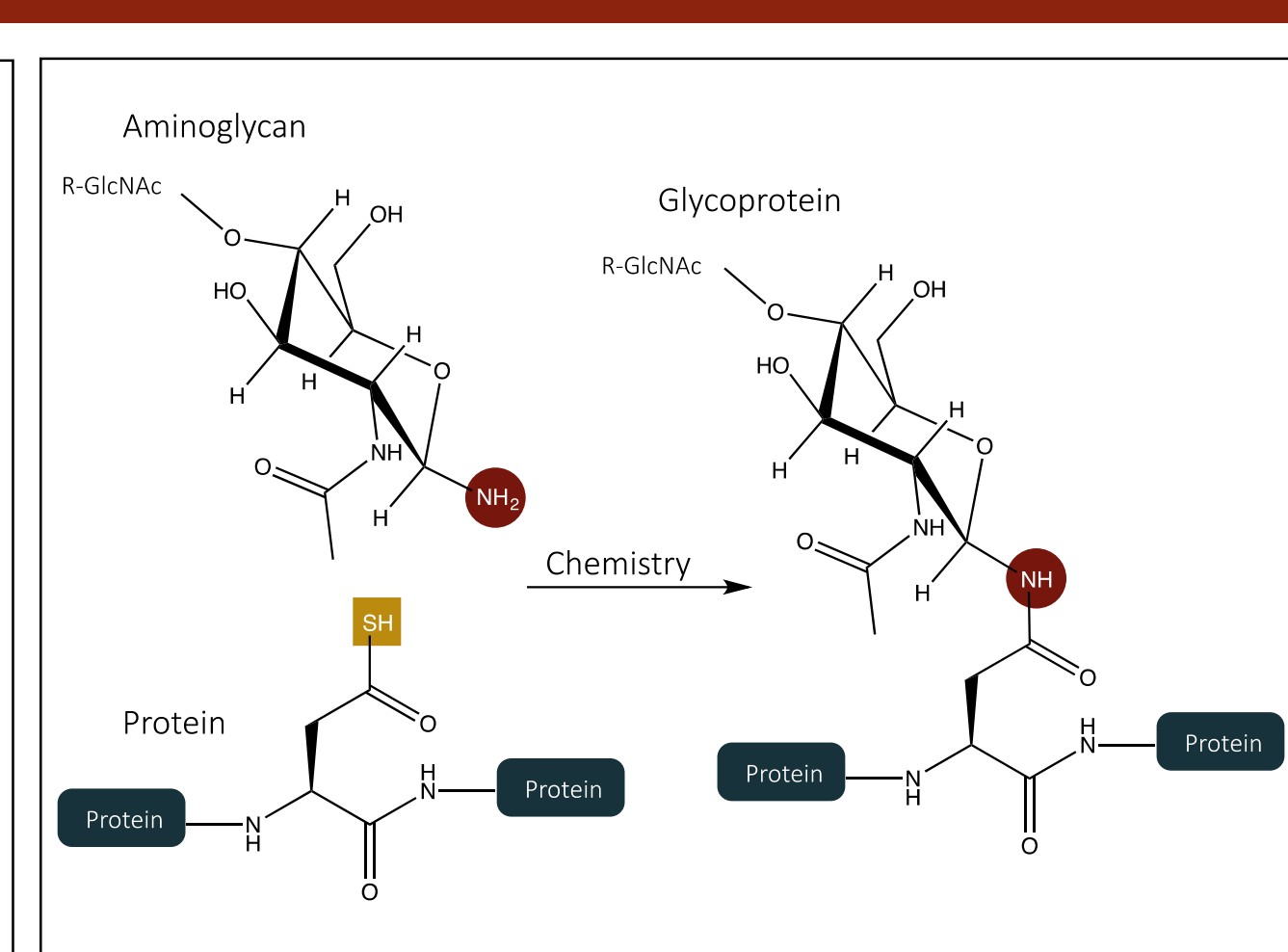
Background

Glycoproteins are important for cellular processes including cell recognition and protein cellular quality control [1]. They are estimated to make up > 50% of proteins [2]. Additionally, they may be the key to unlocking new therapeutic vaccines and cancer immunotherapy treatments [3]. However, glycoproteins remain understudied due to the difficulty of obtaining homogenous samples. A recently published glycoligation method may be a breakthrough if improved [1].

Glycosylation

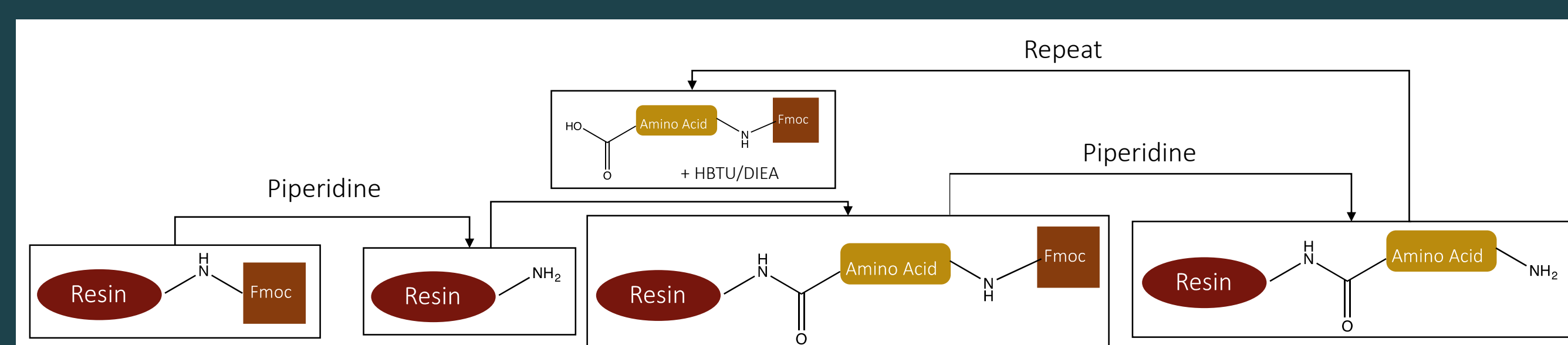


Glycoligation

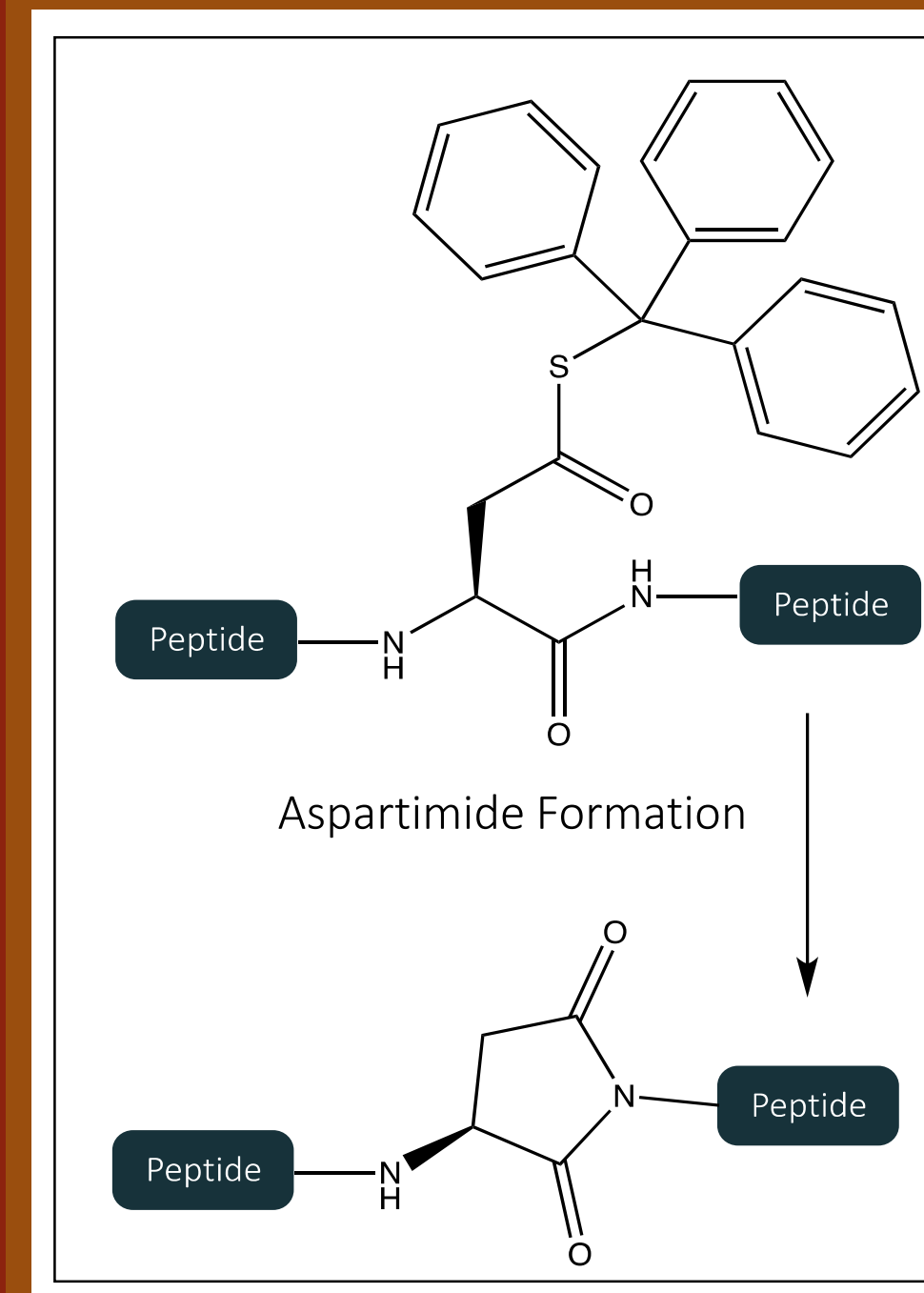


The glycoligation method required peptides to be created using solid phase peptide synthesis using trityl protected aspartic thioester (Asp(S-Trt)) residues at intended ligation sites [4]. However, these thioacid precursors can induce an unwanted aspartimide formation [4]. A method to eliminate the aspartimide formation must be discovered to improve the glycoligation strategy. Aspartyl nitrobenzothiazine (Asp(NBT)) should be resistant to aspartimide formation, and work in place of Asp(S-Trt).

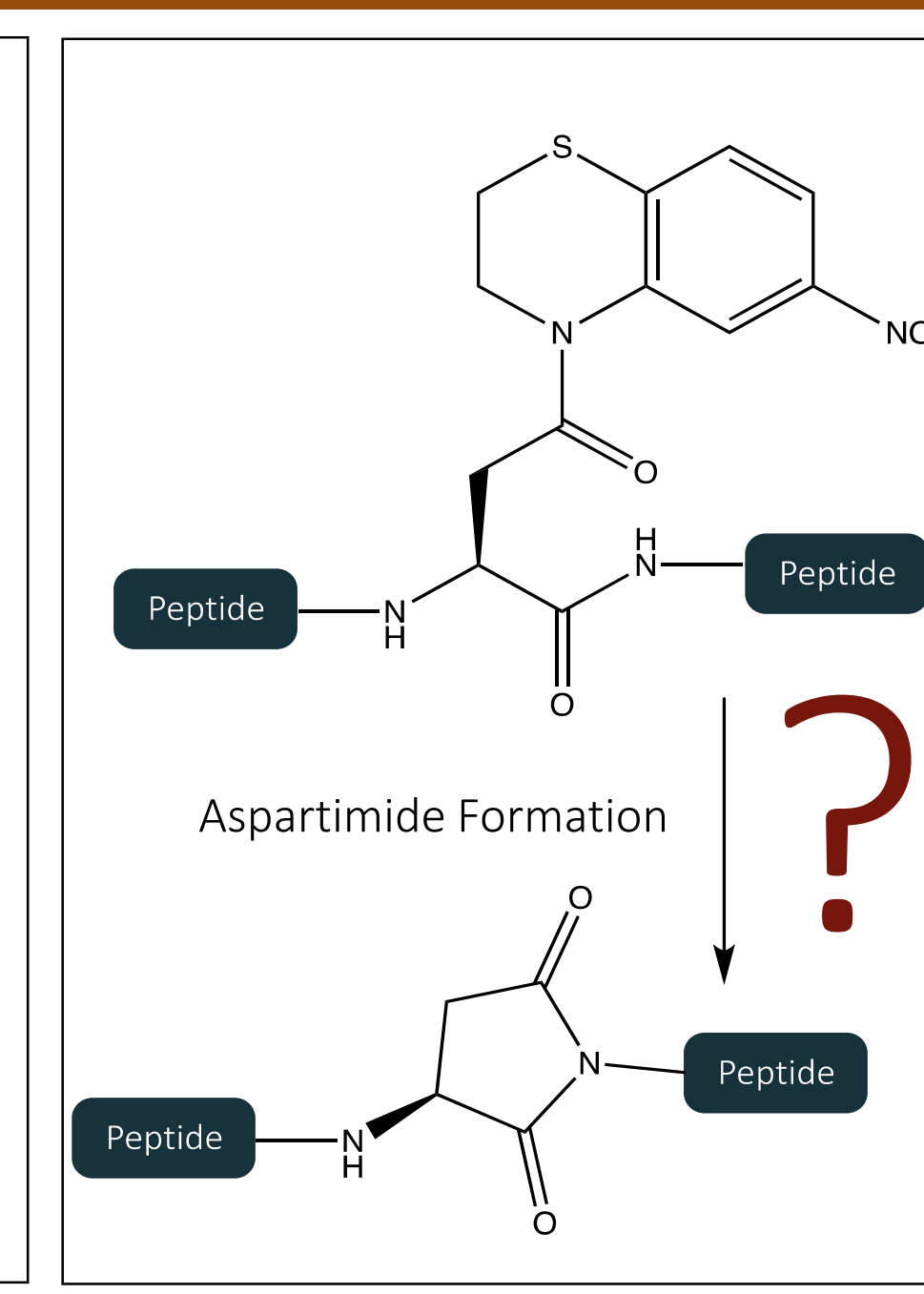
Solid Phase Peptide Synthesis



Asp(S-Trt)

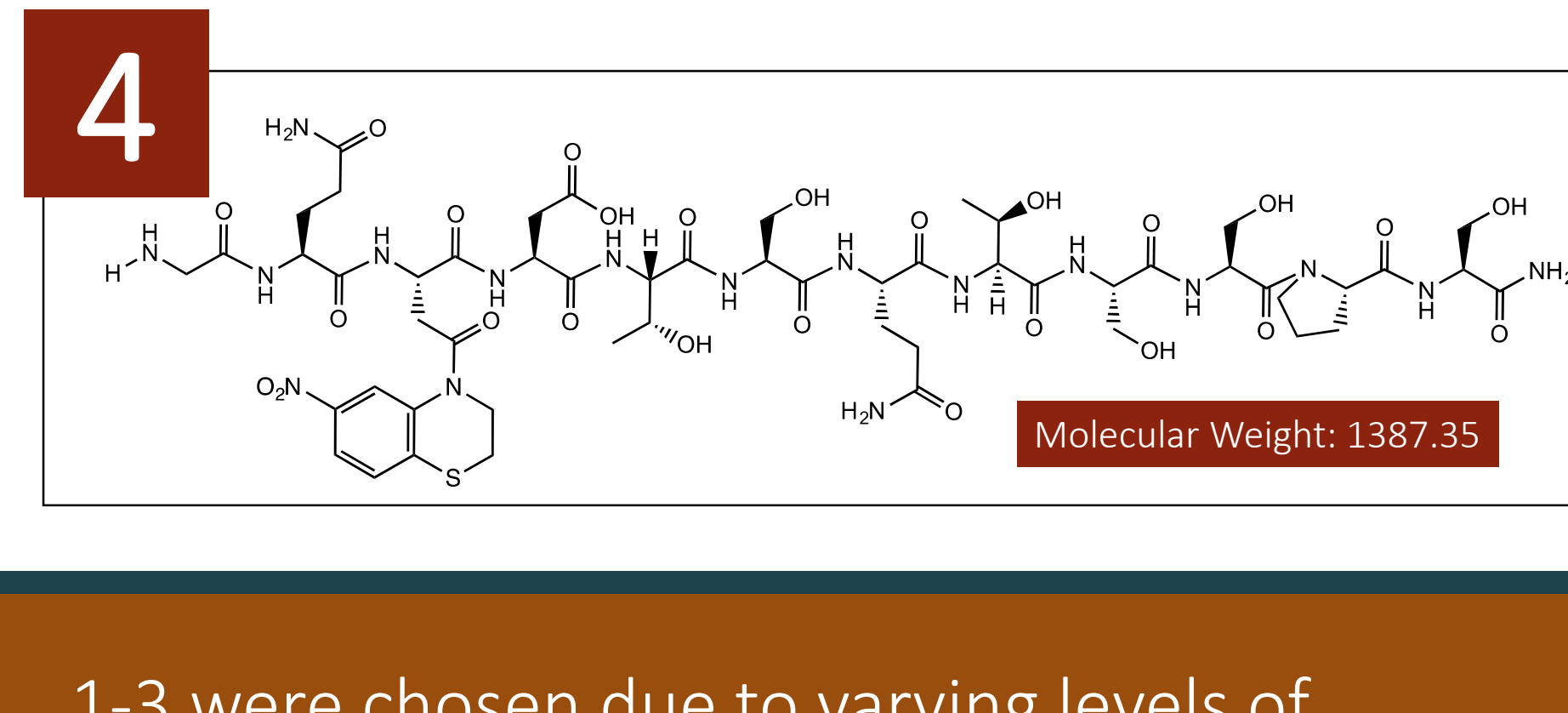
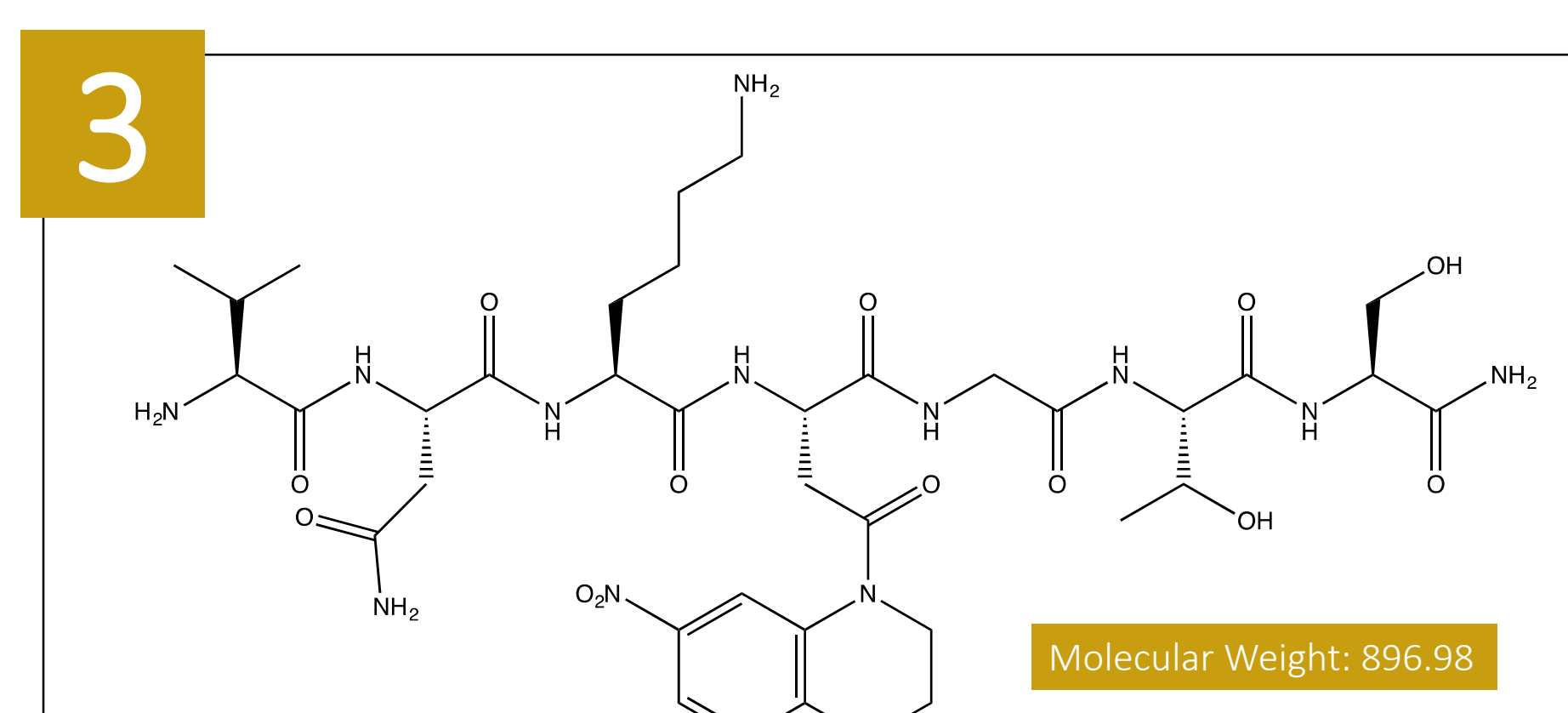
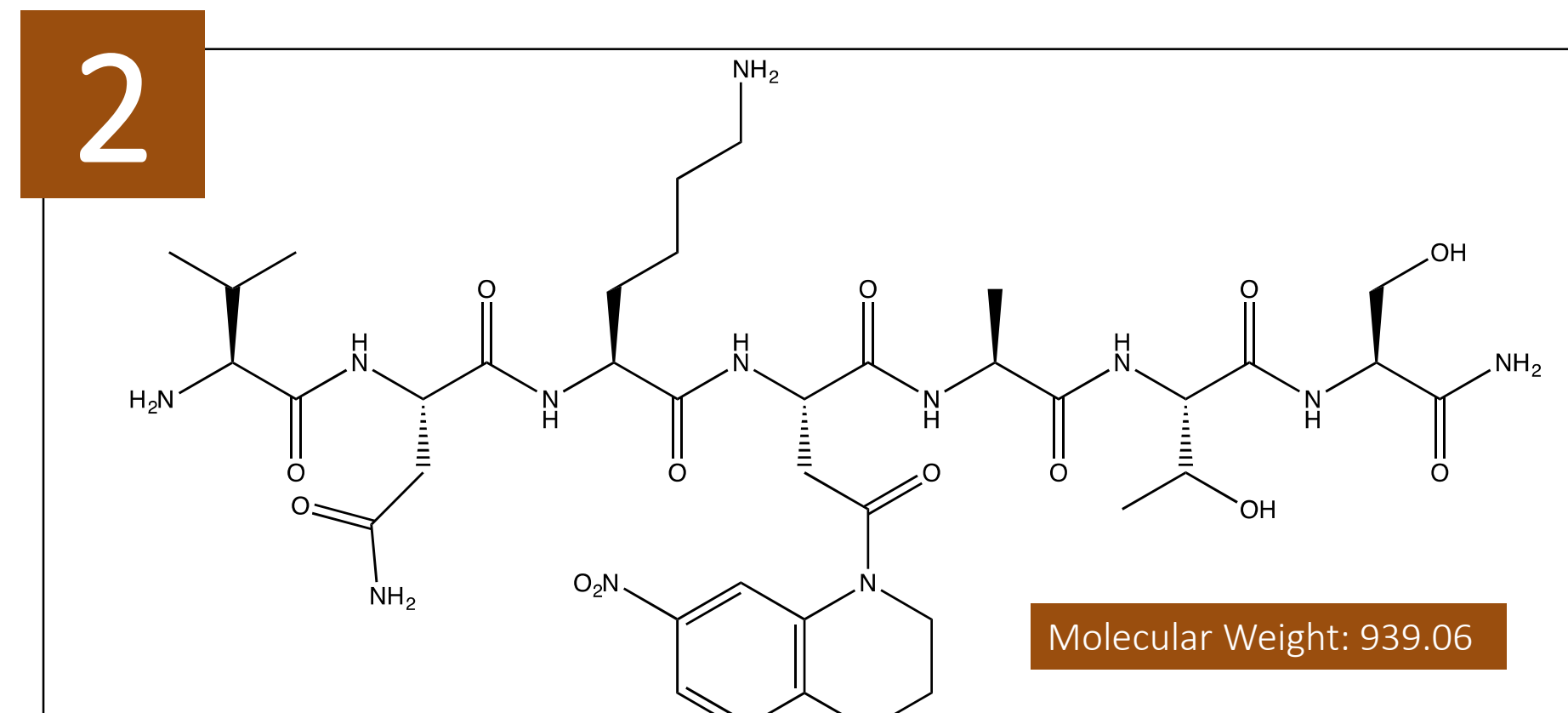
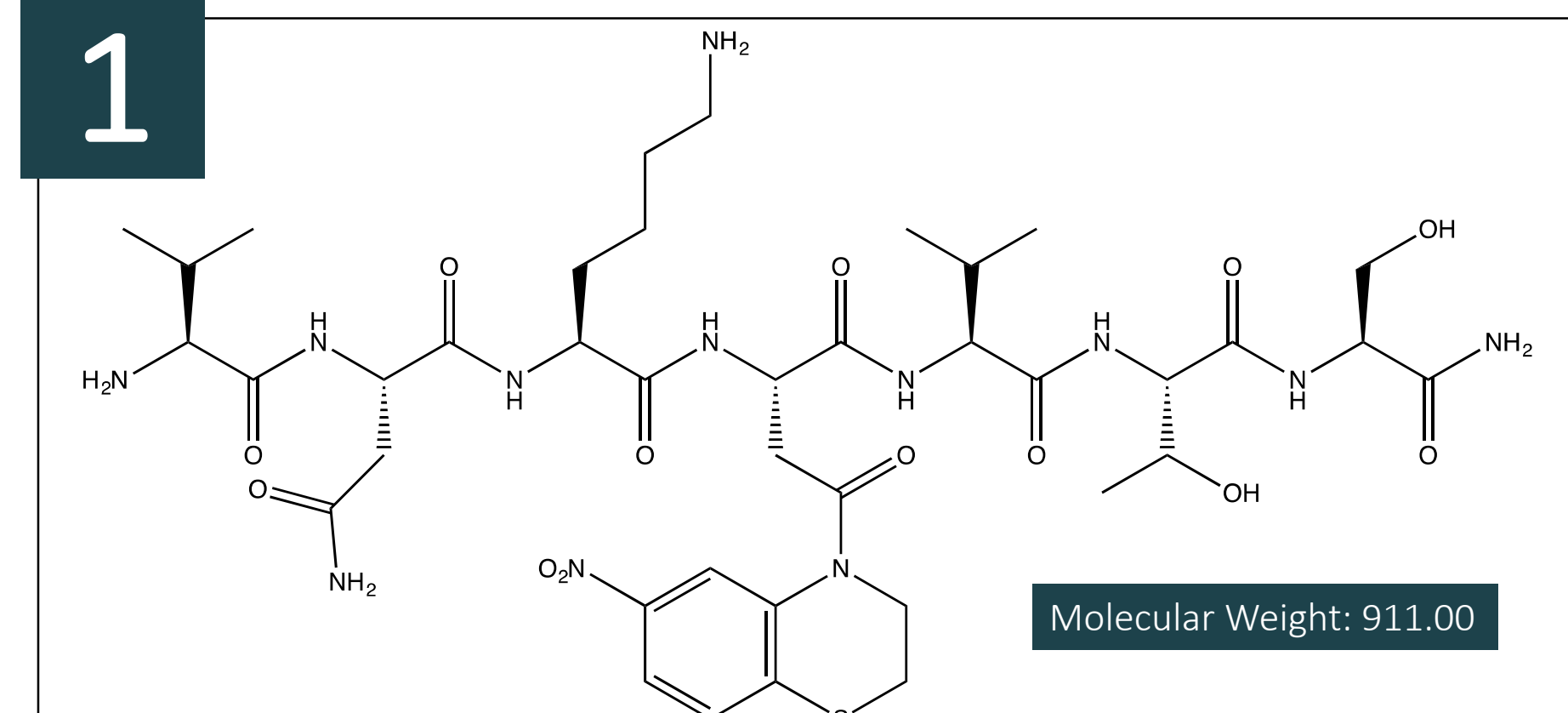


Asp(NBT)



The main objective of this study was producing and analyzing four peptides containing Asp(NBT) to test if Asp(NBT) can work in place of Asp(S-Trt) and avoid aspartimide formation. This was accomplished using the method of Solid Phase Peptide Synthesis [5].

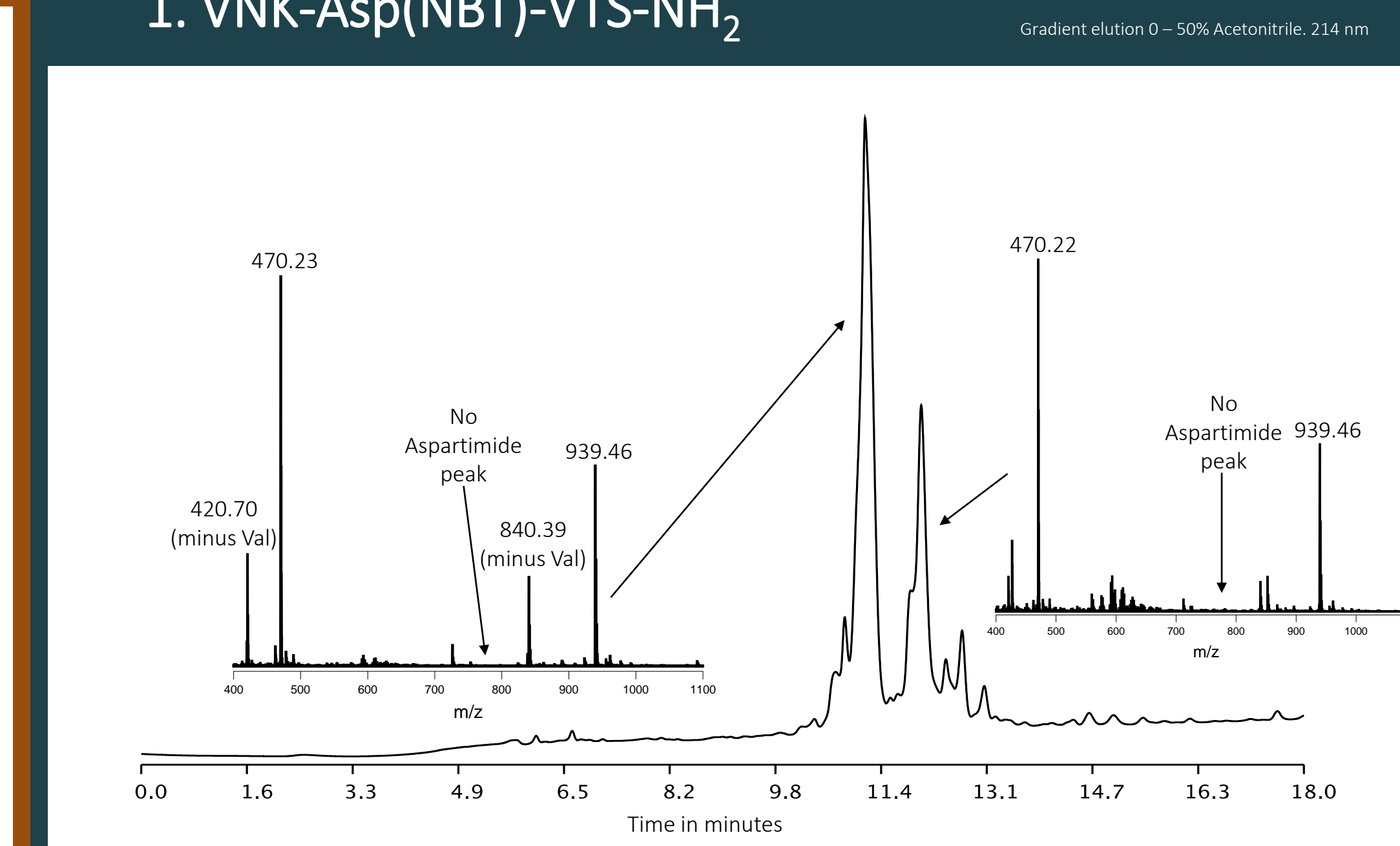
Peptides Produced



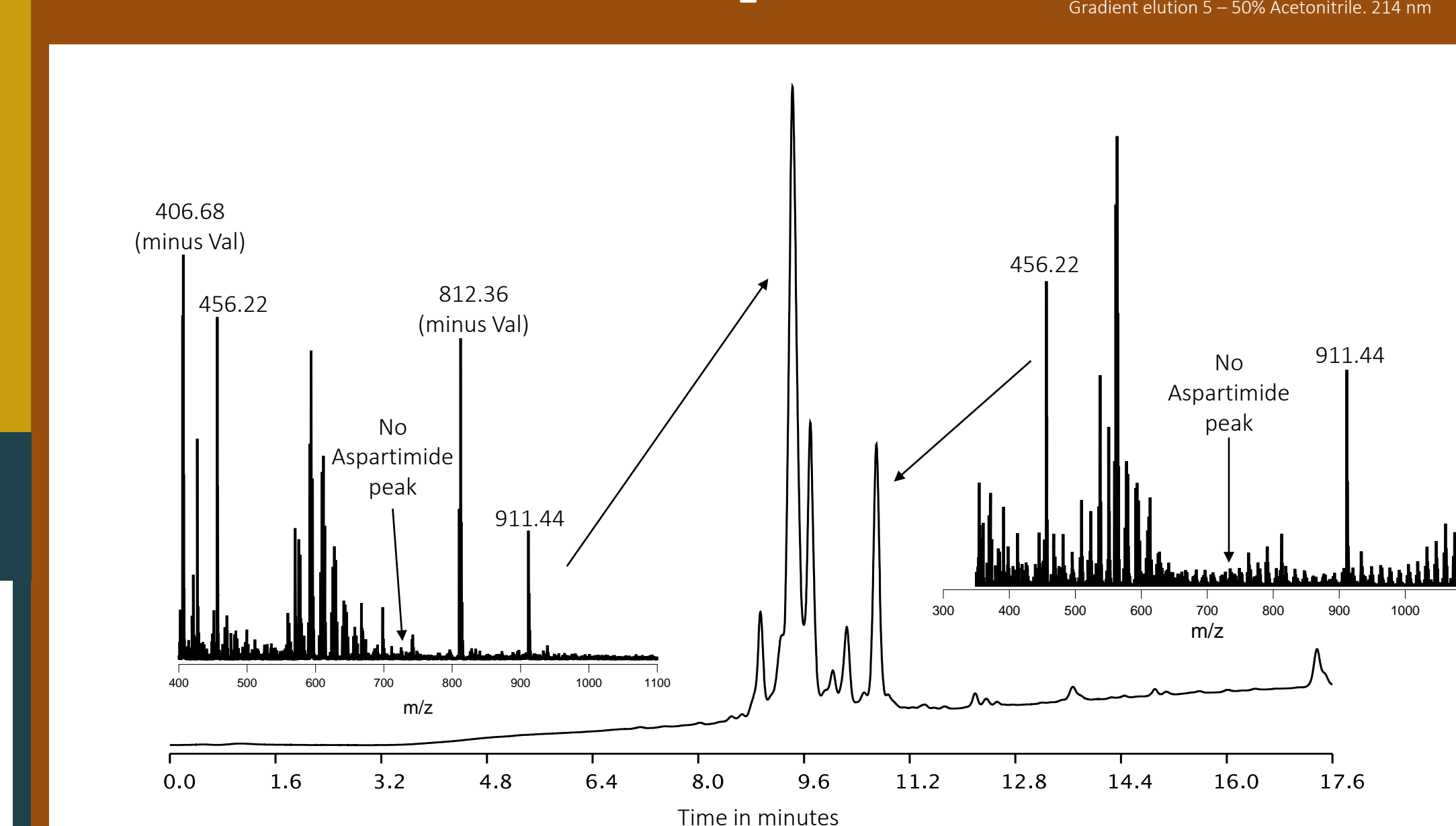
1-3 were chosen due to varying levels of aspartimide formation found using the Asp(S-Trt) versions [4]. 4 was chosen due to its medical significance. The glycosylated and GPI anchored version of 4 is the antigen to the therapeutic antibody Alemtuzumab, a commonly used drug to treat chronic lymphocytic leukemia [6]

Results:

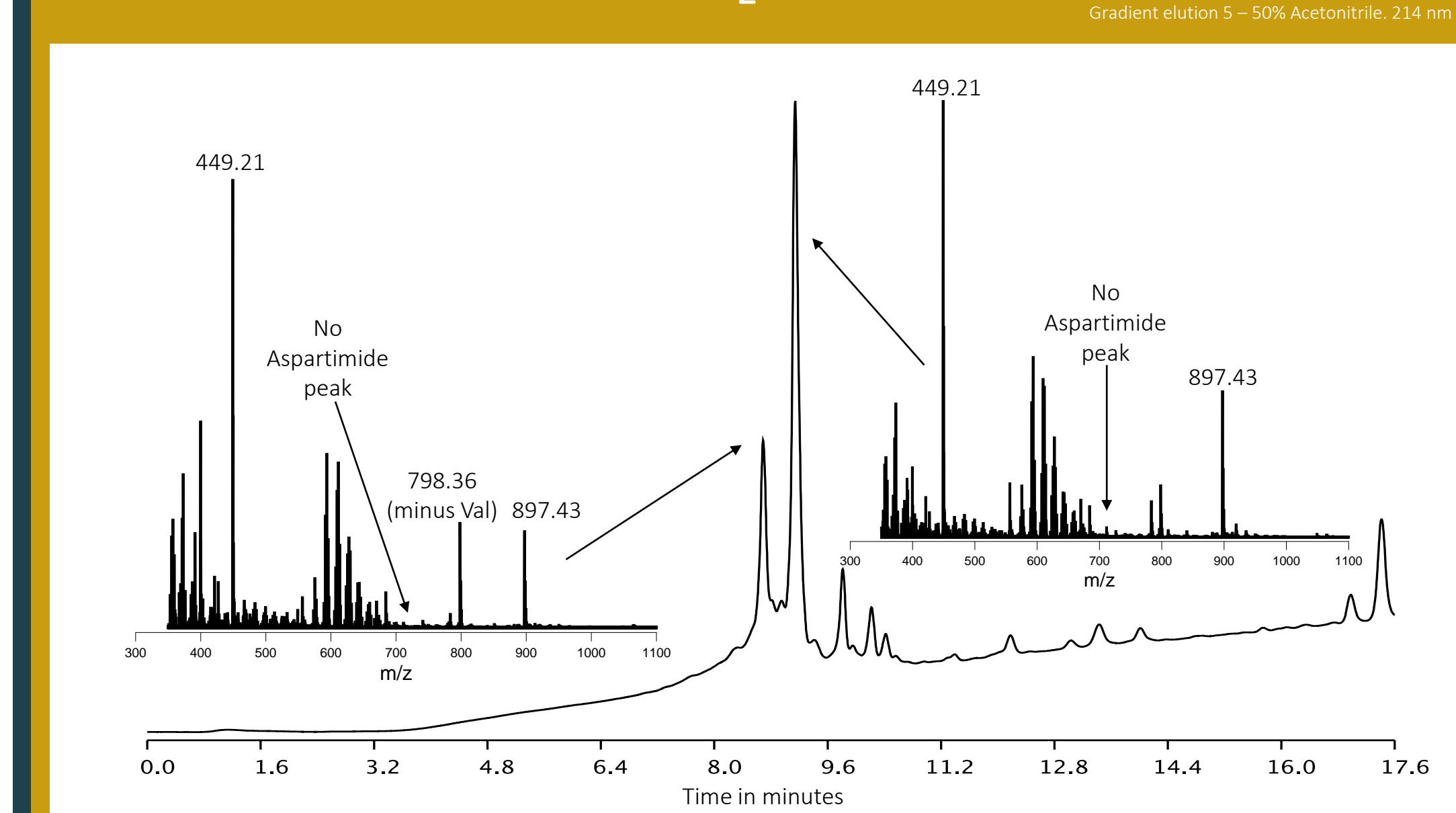
1. VNK-Asp(NBT)-VTS-NH₂



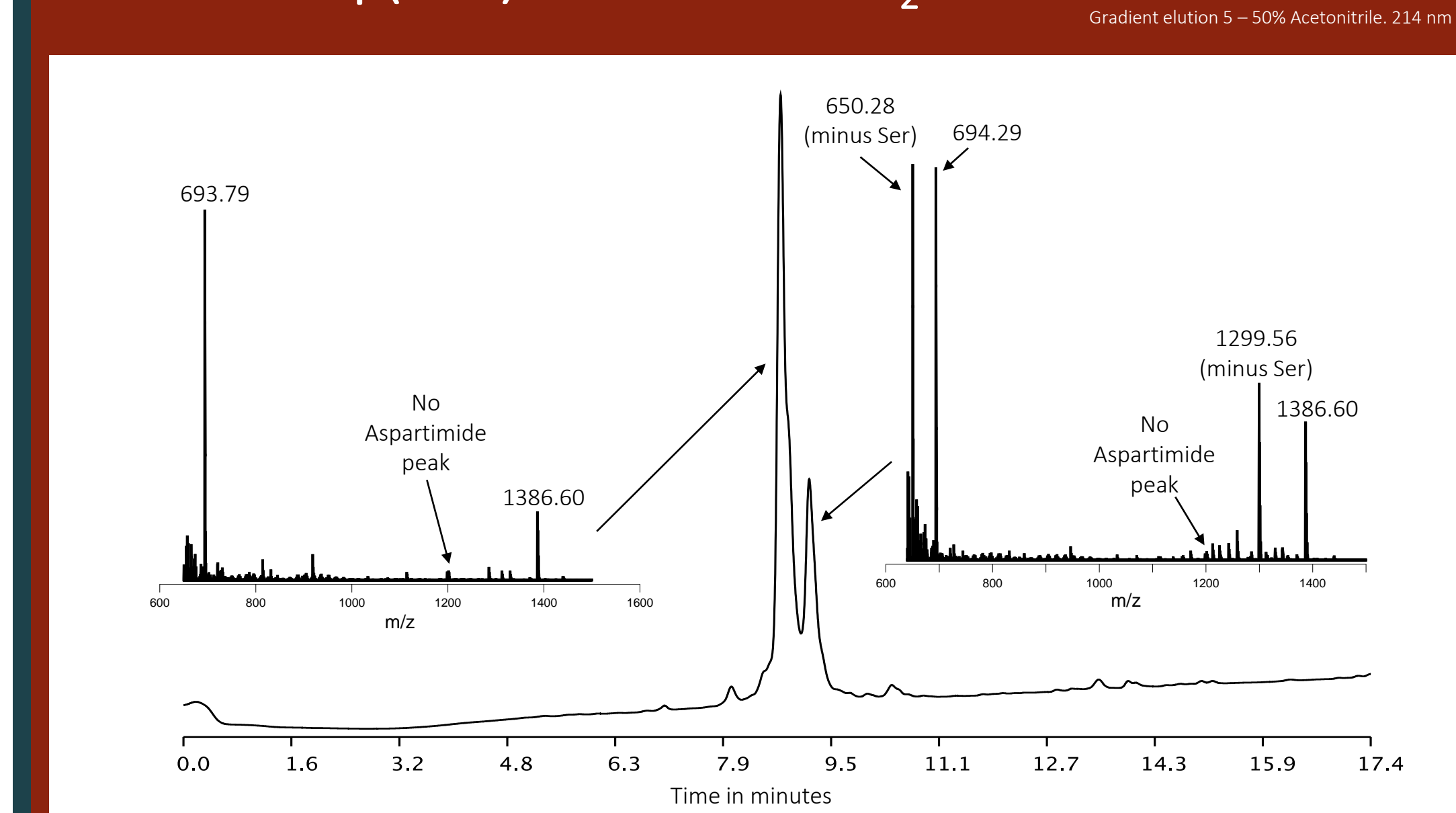
2. VNK-Asp(NBT)-ATS-NH₂



3. VNK-Asp(NBT)-GTS-NH₂



4. GQ-Asp(NBT)-DTSQTSSPS-NH₂



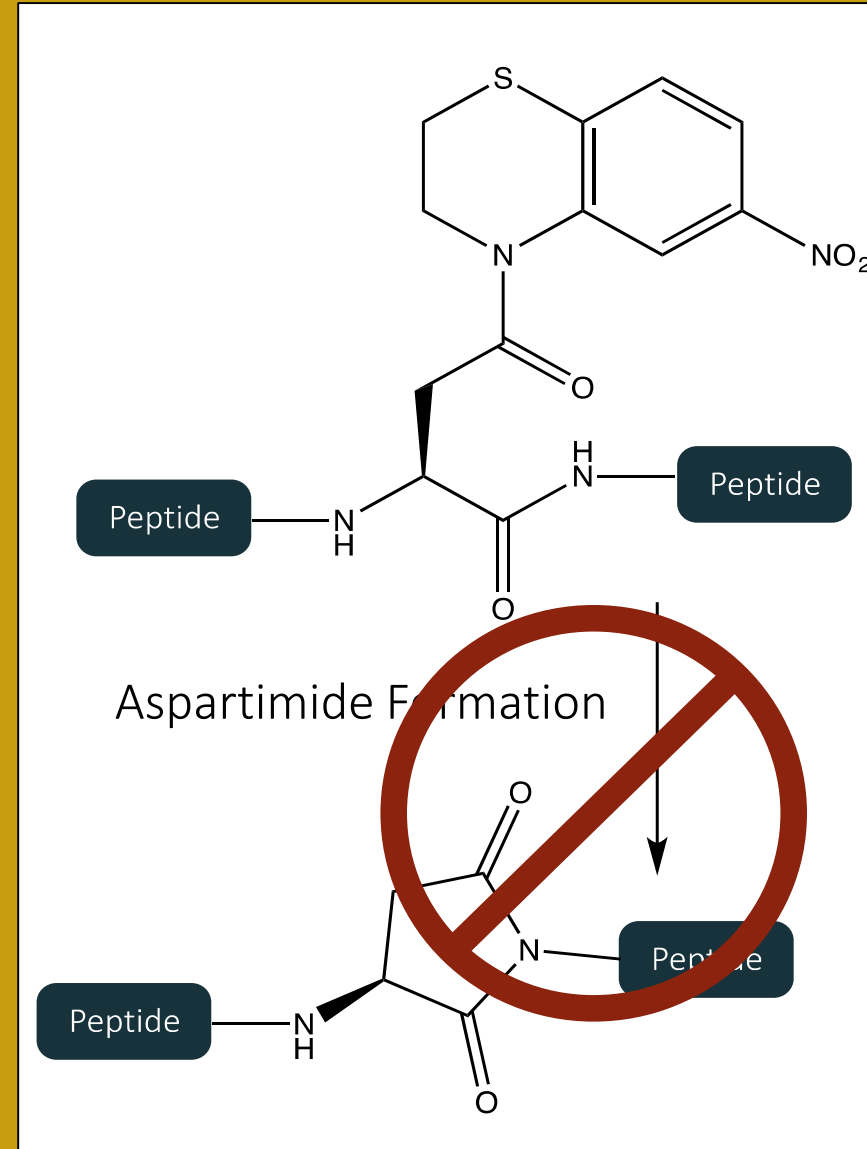
Control peptides were synthesized using aspartate in place of Asp(NBT).

	t _r (min)	mH ⁺ Calc. (g/mol)	mH ⁺ Obs. (g/mol)
1. VNKDVTS-NH ₂	8.24	761.85	761.3
2. VNKDATS-NH ₂	5.04	734.78	733.3
3. VNKDGTS-NH ₂	5.55*	720.75	719.3
4. GQDDTSQTSSPS-NH ₂	6.98	1209.16	1208.9

*100% H₂O isocratic elution. 214 nm

Conclusion

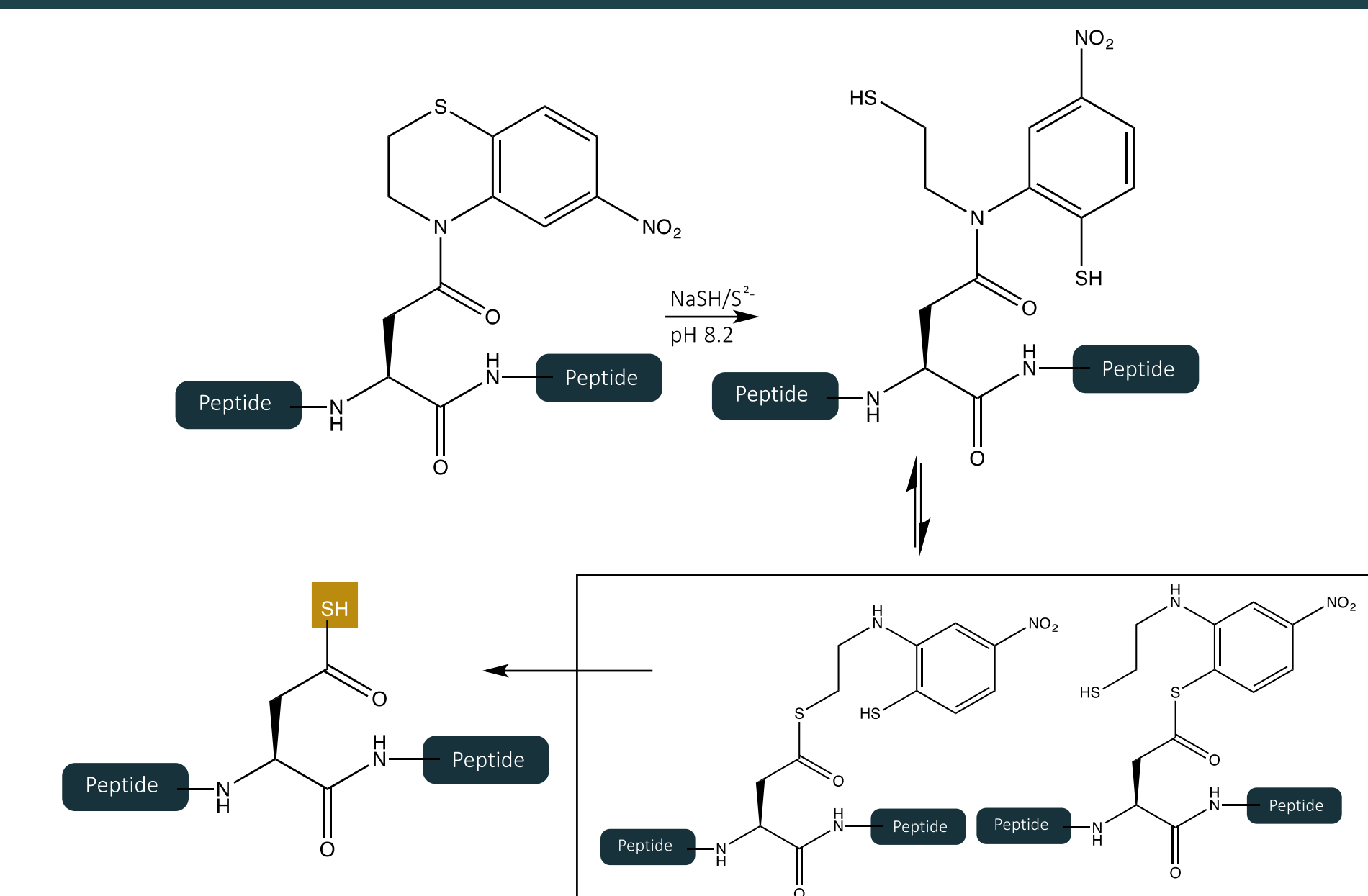
All four Asp(NBT) containing peptides were successfully synthesized without evidence of aspartimide formation. Analysis revealed secondary peaks containing peptide minus Val for 1-3 and peptide minus Ser for 4. These are consistent with control peptides and are likely a product of incomplete coupling.



Next Steps

The peptides synthesized in this study have been sent to the Garner lab at Washington State where the proposed method for turning Asp(NBT) peptide into glycoligation ready aspartic thioacid peptide will be tested. If successful, the amber suppression technique will be used to genetically encode proteins containing Asp(NBT) for use in the first protein glycosylation attempt using the new glycoligation method [7].

Proposed Asp(NBT) Peptide to Aspartic Thioacid Peptide Method



Acknowledgments

The project described was supported by a SURF award provided by the University of Idaho Office of Undergraduate Research.

Other support provided by Peptidaho Research Consortium

Thank you to Gonzaga University for use of the Waters LCT Premier TOF electrospray mass spectrometer funded by NSF CRIF:MU grant #0741868

Thank you to the Odom Corporation as well as Laughing Dog and No Li Breweries for the printing of this poster.

Thank you to the Harbor Center for providing lab space.

D.C. and P.G. work funded by NSF CHE 1149327

- [1] Joseph R, Dyer FB, Garner P (2013) Rapid formation of N-glycopeptides via Cu(II)-promoted glycosylative ligation. *Org Lett* 15:732–735. doi: 10.1021/ol302961s
- [2] Apweiler R, Hermjakob H, Sharon N (1999) On the frequency of protein glycosylation, as deduced from analysis of the SWISS-PROT database. *Biochimica et Biophysica Acta (BBA)*, 1473 (1):4–8.
- [3] Kent SBH (2015) The critical role of peptide chemistry in the life sciences. *J Pept Sci* 21:136–138. doi: 10.1002/psc.2754
- [4] Joseph R, Morales Padilla M, Garner P (2015) Solid phase synthesis of ω-aspartic thioacid containing peptides. *Tetrahedron Lett* 29: 4302-4304. doi: 10.1016/j.tetlet.2015.05.064
- [5] Atherton E, Fox H, Harkiss D, et al (1978) A mild procedure for solid phase peptide synthesis: use of fluorenylmethoxycarbonylamino-acids. *Journal of the Chemical Society, Chemical Communications* 0:537–539. doi: 10.1039/C39780000537
- [6] Lundin J et al. (2002) Phase II trial of subcutaneous anti-CD52 monoclonal antibody alemtuzumab (Campath-1H) as first-line treatment for patients with B-cell chronic lymphocytic leukemia (B-CLL) *Blood* 100:768-773; doi: https://doi.org/10.1182/blood-2002-01-0159
- [7] Liu CC, Schultz PG (2010) Adding new chemistries to the genetic code. *Annu Rev Biochem* 79:413–444. doi: 10.1146/annurev.biochem.052308.105824